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International application number: PCT/US05/005616

International filing date: 23 February 2005 (23.02.2005)

Document type: Certified copy of priority document

Document details: Country/Office: US

Number: 60/547,379

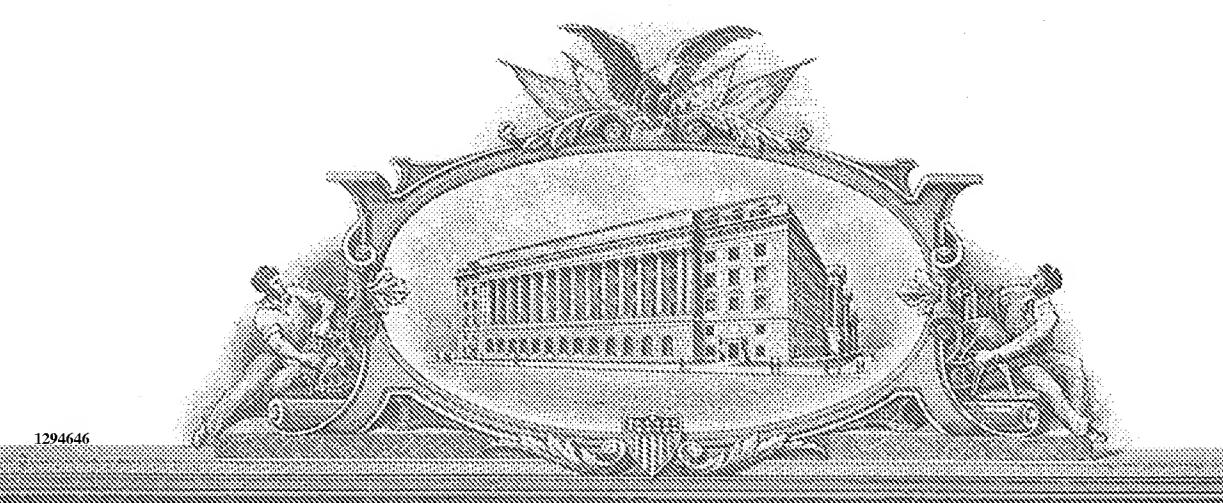
Filing date: 23 February 2004 (23.02.2004)

Date of receipt at the International Bureau: 23 March 2005 (23.03.2005)

Remark: Priority document submitted or transmitted to the International Bureau in

compliance with Rule 17.1(a) or (b)





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APPLICATION NUMBER: 60/547,379
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FILING DATE: February 23, 2004 RELATED PCT APPLICATION NUMBER: PCT/US05/05616

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ENCLOSED APPLICATION PARTS (check all that apply)								
Specification Number of Pages 2 CD(s), Number								
Drawing(s) Number of Sheets Other (specify)								
Application Data Sh	eet. See 37 CFR 1.70	6						
METHOD OF PAYMENT	OF FILING FEES FO	OR THIS PROVISIONAL	. APPLICATION	FOR PATE	NT			
Applicant claims small entity status. See 37 CFR 1.27. FILING FEE								
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This collection of information is required by 37 CFR 1.51. The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 8 hours to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Mail Stop Provisional Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

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PTO/SB/16 (08-03) Approved for use through 07/31/2006. OMB 0651-0032

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Docket Number INVENTOR(S)APPLICANT(S) Residence (City and either State or Foreign Country) Family or Sumame Given Name (first and middle [if any] de CRECY LA JOILA, CA, 92037 EUDES FRANCIS NARIE [Page 2 of 2]

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Number

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Title: Continuous culture apparatus with mobile vessel, allowing selection of fitter cell variants.

SUMMARY

DESCRIPTION

Continuous culture vessels have been used since the 1950's to select for cells that have a higher proliferation rate under given conditions (Monod J., 1950, Ann. Inst. Pasteur. 19:390-410; Novick A., Szilard L., 1950, Science 112:715-716). The power of these devices have never been used to their fullest because they positively select mutants that evade dilution, primarily through attachment to vessel surfaces, resulting in persistent sub-populations of uncontrollable size and growth rate.

One possible solution to overcome this drawback is the implementation of a device with two growth chambers periodically undergoing transient phases of sterilization as described in the patent by Marliere and Mutzel (1999, DE2982162U1).

The solution described in this application is radically different as it consists of using a mobile vessel around the solution instead of moving the solution from one chamber to another.

More specifically, the vessel consists of a sterile flexible tube containing the culture media.

A portion of this tube that can be physically and temporarily separated from the rest of the tube by some lock devices, will contain the cell culture, and will be designated therein as the culture vessel. Fresh medium will be periodically added to the culture by sliding a portion of medium filled tubing.

The design of the lock, see figure, will drive the amount of medium that will be added at each sliding cycle.

Spend culture and medium will also be eliminated through a tubing sliding step.

The frequency of the movement of the tube can be set by the experimenter reproducing a chemostat regime or can be for example regulated through a feedback loop depending on the turbidity of the culture in the vessel that will be read via a turbidometer, emulating a turbidostat regime. Other regulation approaches can be implemented.

The system is designed to allow the complete rotation of the vessel and the locks.

The positioning of the vessel will be adjusted in accordance to the aeration cycle.

Aeration can be provided two ways:

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- 1- by passive diffusion of ambient air through gas permeable tubing,
- 2- by injecting filtered air that will first bubble through the media and then be introduced in the vessel through the entrance locking sub-system.

Previously the spend air will have been eliminated by rotation of the vessel in the upright position as described in attached figures.

The presence of several gates (superior to 2) should avoid any contamination in the tubing of the medium in the up chamber tubing. However several UV gates can be added upstream and downstream of the culture vessel for additional security.

Inoculation will be performed by sterily injecting a cell culture through the lower lock that would have been set in the opened position.

An added lock will eventually be added downstream of the chamber lock to create a sampling chamber.

Tubing specifications: The tubing must be large enough to avoid capillarity problems that would prevent the air from circulating freely. The tubing should sustain sterilization cycle, be gas permeable, be flexible enough to be closed by compression and be clear enough to allow any optical measurement.

